



Human Pancreatic Microvascular Endothelial Cells (HPaMEC) Catalog #3800

Cell Specification

The pancreas is an endocrine gland and digestive organ which secretes hormones and produces pancreatic juice to aid in digestion. Pancreatic endocrine cells, also known as pancreatic islets, produce vital hormones such as glucagon, insulin, amylin, and somatostatin. In order to support the islets, the pancreas is a highly vascularized organ where endothelial cells play a critical role [1, 2]. Pancreatic microvascular endothelial cells (PaMEC) not only function to transport oxygen and nutrients to the pancreas, but also affect beta cell function and proliferation, influence insulin gene expression during islet development, and produce various growth factors associated with vasoactivity [1, 2]. Additionally, PaMEC are involved in optimizing blood glucose sensing and regulation [1, 2]. Recent studies have shown that PaMEC are involved in the development and progression of pancreatic cancer [3]. Human PaMEC (HPaMEC) are a useful *in vitro* model for studying islet biology, pancreatic cancer, transplantation, and regenerative medicine.

HPaMEC from ScienCell Research Laboratories are isolated from human pancreas. HPaMEC are cryopreserved at passage one and delivered frozen. Each vial contains $>5 \times 10^5$ cells in 1 ml volume. HPaMEC are characterized by immunofluorescence with antibodies specific to vWF/Factor VIII and CD31 (PECAM). HPaMEC are negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast, and fungi. HPaMEC are guaranteed to further expand for 10 population doublings under the conditions provided by ScienCell Research Laboratories.

Recommended Medium

It is recommended to use Endothelial Cell Medium (ECM, Cat. #1001) for culturing HPaMEC *in vitro*.

Product Use

HPaMEC are for research use only. They are not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

Storage

Upon receiving, directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

Shipping

Dry ice.

References

- [1] Zanone M, Favaro E, Camussi G. (2008) "From endothelial to beta cells: insights into pancreatic islet microendothelium." *Curr Diabetes Rev.* 4(1): 1-9.
- [2] Konstantinova I and Lammert E. (2004) "Microvascular development: learning from pancreatic islets." *Bioessays* 26(10): 1069-1075.
- [3] Nielsen M, Mortensen M, Detlefsen S. (2016) "Key players in pancreatic cancer stroma interaction: cancer-associated fibroblasts, endothelial and inflammatory cells." *World J Gastroenterol* 22(9): 2678-2700.

Instructions for culturing primary cells

Caution: Cryopreserved primary cells are very delicate. Thaw the vial in a 37°C water bath and return the cells to culture as quickly as possible with minimal handling! Do not centrifuge the cells after thawing as this can damage the cells.

Initiating the culture:

Note: ScienCell primary cells must be cultured in a 37°C, 5% CO₂ incubator. Cells are only warranted if ScienCell media and reagents are used and the recommended protocols are followed.

1. Prepare a fibronectin-coated culture vessel (2 µg/cm², T-75 flask is recommended). To obtain a 2 µg/cm² fibronectin-coated culture vessel, add 5 ml of sterile Dulbecco's phosphate buffered saline, Ca⁺⁺- and Mg⁺⁺-free (Cat. #0303) to a T-75 flask and then add 150 µl of fibronectin stock solution (Cat. #8248). Leave the vessel in a 37°C incubator overnight (or for at least 2 hours).
2. Prepare complete medium. Decontaminate the external surfaces of medium bottle and medium supplement tubes with 70% ethanol and transfer them to a sterile field. Aseptically transfer supplement to the basal medium with a pipette. Rinse the supplement tube with medium to recover the entire volume.
3. Aspirate the fibronectin solution and add 15 ml of complete medium to the culture vessel. The fibronectin solution can be used twice. Leave the vessel in the sterile field and proceed to thaw the cryopreserved cells.
4. Place the frozen vial in a 37°C water bath. Hold and rotate the vial gently until the contents completely thaw. Promptly remove the vial from the water bath, wipe it down with 70% ethanol, and transfer it to the sterile field.
5. Carefully remove the cap without touching the interior threads. Gently resuspend and dispense the contents of the vial into the equilibrated, fibronectin-coated culture vessel.

Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to the cells than the effect of residual DMSO in the culture. It is also important that cells are plated in fibronectin-coated culture vessels to promote cell attachment.

6. Replace the cap or lid of the culture vessel and gently rock the vessel to distribute the cells evenly. Loosen cap, if necessary, to allow gas exchange.
7. Return the culture vessel to the incubator.
8. Do not disturb the culture for at least 16 hours after the culture has been initiated. Refresh culture medium the next day to remove residual DMSO and unattached cells.

Maintaining the culture:

1. Refresh supplemented culture medium the next morning after establishing a culture from cryopreserved cells.
2. Change the medium every three days thereafter, until the culture is approximately 70% confluent.

3. Once the culture reaches 70% confluency, change medium every other day until the culture is approximately 90% confluent.

Subculturing:

1. Subculture when the culture reaches 90% confluency.
2. Prepare fibronectin-coated culture vessels ($2 \mu\text{g}/\text{cm}^2$) one day before subculture.
3. Warm complete medium, trypsin/EDTA solution, 0.05% (T/E, Cat. #0183), T/E neutralization solution (TNS, Cat. #0113), and DPBS (Ca^{++} - and Mg^{++} -free, Cat. #0303) to **room temperature**. We do not recommend warming reagents and medium in a 37°C water bath prior to use.
4. Rinse the cells with DPBS.
5. Add 8 ml DPBS and 2 ml 0.05% T/E solution (Cat. #0183) into flask (in the case of a T-75 flask). Gently rock the flask to ensure complete coverage of cells by T/E solution. Incubate the flask at room temperature until most cells completely round up (typically between 1-5 minutes). Use a microscope to monitor the change in cell morphology.

Note: We recommend using ScienCell 0.05% T/E solution which is optimized to minimize cell damage due to over trypsinization. If 0.25% T/E solution (Cat. #0103) is used, then 10 ml DPBS and 400 μl 0.25% T/E solution should be used.

6. During incubation, prepare a 50 ml conical centrifuge tube with 5 ml of fetal bovine serum (FBS, Cat. #0500).
7. Transfer T/E solution from the flask to the 50 ml centrifuge tube (a small percent of cells may detach) and continue to incubate the flask at 37°C for another minute (no solution in the flask).
8. At the end of incubation, gently tap the side of the flask to dislodge cells from the surface. Check under a microscope to make sure that all cells detach.
9. Add 5 ml of TNS solution to the flask and transfer detached cells to the 50 ml centrifuge tube. Rinse the flask with another 5 ml of TNS to collect the residual cells.
10. Examine the flask under a microscope for a successful cell harvest by looking at the number of cells being left behind; there should be less than 5%.
11. Centrifuge the 50 ml tube at 1000 rpm for 5 minutes. Resuspend cells in culture medium.
12. Count and plate cells in a new fibronectin-coated culture vessel with the recommended cell density. A seeding density of 5,000-7,000 cells/ cm^2 is recommended.

Note: We do not recommend cryopreservation of primary cells by the end user. Refreezing cells may damage them and affect cell performance. ScienCell does not guarantee primary cells cryopreserved by the end user.

Caution: Handling human derived products is potentially biohazardous. Although each cell strain tests negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate, therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working with these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].

[1] Grizzle WE, Polt S. (1988) "Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues." *J Tissue Cult Methods*. 11: 191-9.