Relative Human Telomere Length Quantification qPCR Assay Kit (RHTLQ)

Catalog #8908
100 reactions

Product Description

Telomeres are repetitive nucleotide elements at the ends of chromosomes that protect chromosomes from degradation and genetic information loss. Normal diploid cells lose telomeres with each cell cycle. Telomere length, therefore, decreases over time and may predict lifespan. Telomere shortening has negative effects on health conditions and has been linked to many health issues including aging and cancer. Accurate and consistent quantification of telomere length is important in many aspects of cell biology such as chromosomal instability, DNA repair, senescence, apoptosis, cell dysfunctions, and oncogenesis.

ScienCell's Relative Human Telomere Length Quantification qPCR Assay Kit (RHTLQ) is designed to directly compare the average telomere length of the samples. The telomere primer set recognizes and amplifies telomere sequences. The single copy reference (SCR) primer set recognizes and amplifies a 100 bp-long region on human chromosome 17, and serves as reference for data normalization. The carefully designed primers ensure: (i) high efficiency for trustworthy quantification; and (ii) no non-specific amplification. Each primer set has been validated by qPCR with melt curve analysis and gel electrophoresis for amplification specificity and by template serial dilution for amplification efficiency.

Kit Components

Cat #	Component	Quantity	Storage
8908a	Telomere primer set, lyophilized	1 vial	-20°C
8908b	Single copy reference (SCR) primer set, lyophilized	1 vial	-20°C
8908c	Nuclease-free H_2O	4 mL	4°C

Additional Materials Required (Materials Not Included in Kit)

Component	Recommended
genomic DNA template	Customers' samples
qPCR plate or tube	
qPCR master mix	FastStart Essential DNA Green Master (Roche, Cat #06402712001)

Quality Control

The specificity of the primer sets are validated by qPCR with melt curve analysis. The PCR products are analyzed by gel electrophoresis. The efficiency of the primer sets are validated by template serial dilution (See **Appendix**).

Product Use

RHTLQ is for research use only. It is not approved for human or animal use, or for application in clinical or *in vitro* diagnostic procedures.

Shipping and Storage

The product is shipped at ambient temperature. Upon receipt, store the primers at -20°C in a manual defrost freezer, and nuclease-free H_2O at 4°C.

Procedures

Important:* Only** *use polymerases with hot-start capability to prevent possible primer-dimer formation.* ***Only *use nuclease-free reagents in PCR amplification.*

Note: RHTLQ is optimized using FastStart Essential DNA Green Master (Roche, Cat #06402712001) and is highly recommended. Use of other qPCR master mixes may compromise results.

1. Prior to use, allow vials (Cat #8908a and #8908b) to warm to room temperature.
2. Centrifuge the vials at 1,500x g for 1 minute.
3. Add 200 µl nuclease-free H_2O (Cat #8908c) to telomere primer set (lyophilized, Cat #8908a) to make telomere primer stock solution. Aliquot as needed. Store at -20°C in a manual defrost freezer. Avoid repeated freeze-and-thaw cycles.
4. Add 200 µl nuclease-free H2O (Cat #8908c) to SCR primer set (lyophilized, Cat #8908b) to make SCR primer stock solution. Aliquot as needed. Store at -20°C in a manual defrost freezer. Avoid repeated freeze-and-thaw cycles.
5. For each genomic DNA sample, prepare two qPCR reactions, one with telomere primer stock solution, and one with SCR primer stock solution. Prepare 20 µl qPCR reactions for one well as shown in Table 1.

Table 1.

Genomic DNA template	0.5 – 20 ng
Primer stock solution (Telomere or SCR)	2 µl
2x qPCR master mix	10 µl
Nuclease-free H_2O (Cat #8908c)	variable
Total volume	***20 µl***

6. Seal the qPCR reaction wells. Centrifuge the plates or tubes at 1,500x g for 15 seconds. For maximum reliability, replicates are strongly recommended (minimum of 3).
7. For qPCR program setup, please refer to the instructions of the master mix of the user's choice. We recommend a typical 3-step qPCR protocol as shown in Table 2:

Table 2.

Step	Temperature	Time	Number of cycles
Initial denaturation	95°C	10 min	1
Denaturation	95°C	20 sec	32
Annealing	52°C	20 sec	
Extension	72°C	45 sec	
Data acquisition	Plate read		
Optional	*Melting curve analysis*		1
Hold	20°C	Indefinite	1

Figure 1. A typical amplification curve showing the amplification of a qPCR product.

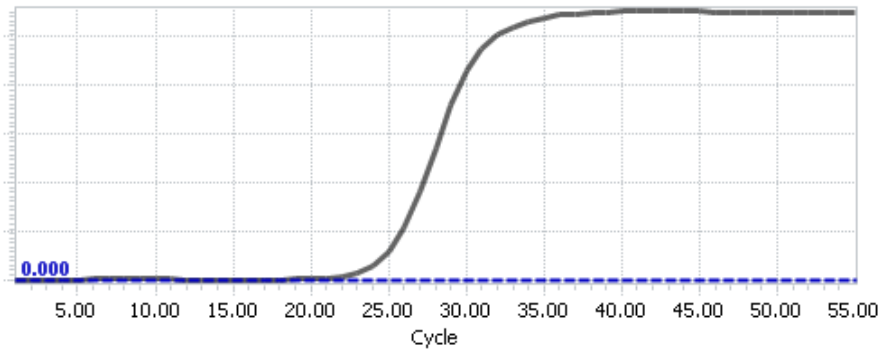
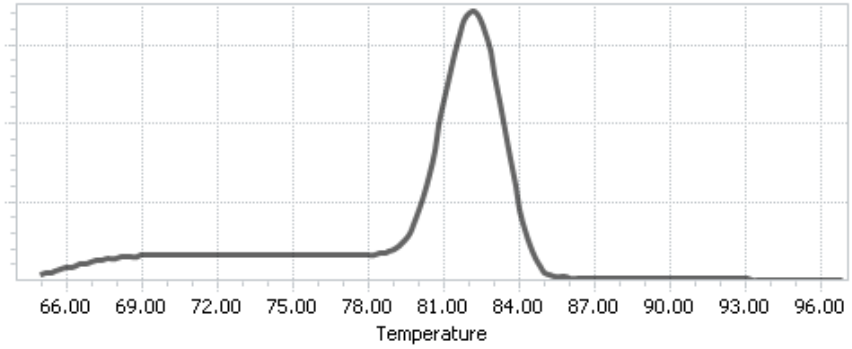


Figure 2. A typical melting peak of a qPCR product.



Quantification Method: Comparative ΔΔCq (Quantification Cycle Value) Method

Note: Please refer to your qPCR instrument's data analysis software for data analysis. The method provided here serves as guidance for quick manual calculations.

1. For telomere (TEL), ΔCq (TEL) is the quantification cycle number difference of TEL between two genomic DNA samples.

   ΔCq (TEL) = Cq (TEL, sample 2) - Cq (TEL, sample 1)

   Note: the value of ΔCq (TEL) can be positive, 0, or negative.

2. For single copy reference (SCR), ΔCq (SCR) is the quantification cycle number difference of SCR between two genomic DNA samples.

   ΔCq (SCR) = Cq (SCR, sample 2) - Cq (SCR, sample 1)

   Note: the value of ΔCq (SCR) can be positive, 0, or negative.

3. ΔΔCq = ΔCq (TEL) - ΔCq (SCR)

4. Relative telomere length of sample 2 to sample 1 (fold) = $2^{-\Delta\Delta Cq}$

Example Calculations: Comparative ΔΔCq (Quantification Cycle Value) Method

Table 3. Cq (Quantification Cycle) values of telomere qPCR (TEL) and single copy reference qPCR (SCR) obtained for two genomic DNA samples.

Primer set	*Sample 1*	*Sample 2*
TEL	16.84	14.16
SCR	26.43	25.20

ΔCq (SCR) = Cq (SCR, sample 2) - Cq (SCR, sample 1)
= 25.20 - 26.43
= -1.23

ΔCq (TEL) = Cq (TEL, sample 2) - Cq (TEL, sample 1)
= 14.16 - 16.84
= -2.68

ΔΔCq = ΔCq (TEL) - ΔCq (SCR)
= -2.68- (-1.23)
= -1.45

$$\text{Relative telomere length of sample 2 to sample 1 (fold)} = 2^{-\Delta\Delta Cq}$$
$$= 2^{1.45}$$
$$= 2.73$$

Example Conclusions: The average telomere length of sample 2 is 2.73 fold longer than that of sample 1.

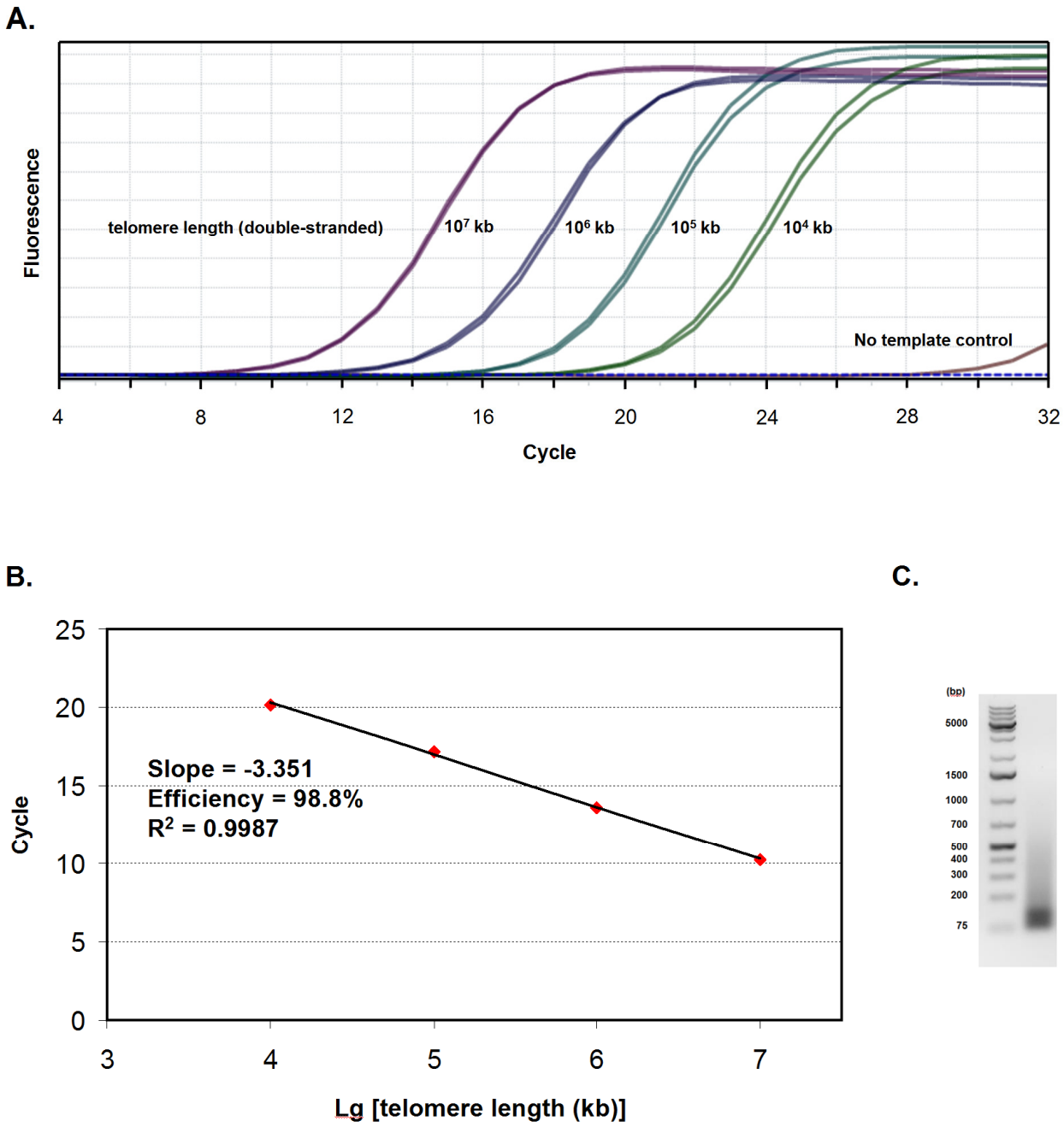


Figure 3. Quality assessment of Telomere primer set. **(A)** qPCR amplification curves using serially diluted telomere repeats as template. **(B)** Derivation of qPCR efficiency of Telomere primer set. **(C)** Separation of Telomere qPCR product by gel electrophoresis. A smeared band is observed as expected.

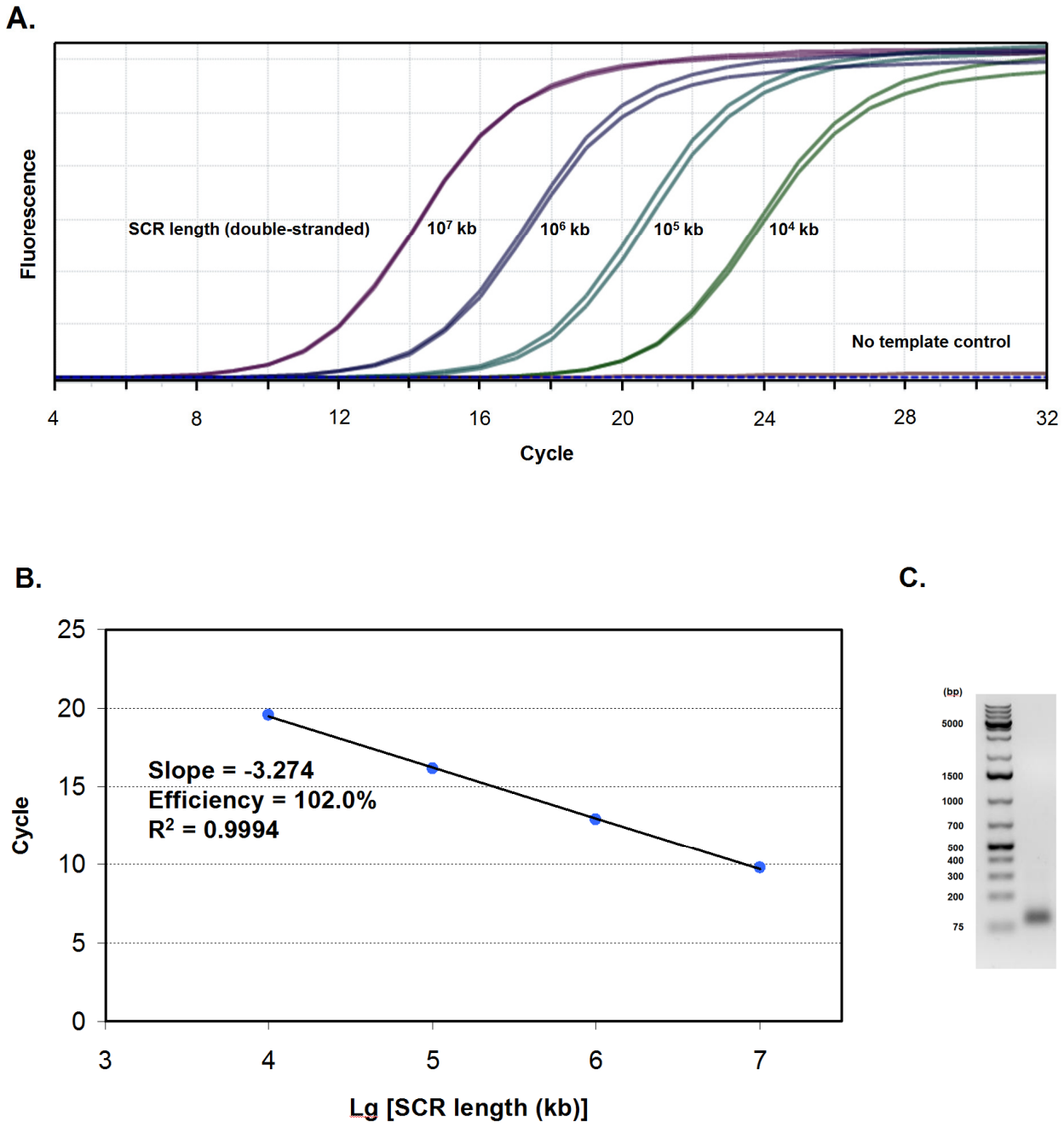


Figure 4. Quality assessment of Single copy reference (SCR) primer set. **(A)** qPCR amplification curves using serially diluted SCR template. **(B)** Derivation of qPCR efficiency of SCR primer set. **(C)** Separation of SCR qPCR product by gel electrophoresis.